

## STRUCTURAL–FUNCTIONAL ANALYSIS OF BIOPOLYMERS AND THEIR COMPLEXES

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# Role of the Structural Context in Selection of Hydrophobic Side-Chain Rotamers in a- and d-Positions of $\alpha$ -Helices

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**Abstract**—It was demonstrated for the first time that the distribution of side-chain rotamers in the a- and d-positions of  $\alpha$ -helices of coiled-coil (cc) proteins follows a certain trend, rather than being random. For instance, most side chains adopt t rotamers in the a-positions and  $g^-$  rotamers in the d-positions of helical dimers. Vice versa, most side chains adopt  $g^-$  rotamers in the a-positions and t rotamers in the d-positions of tetramers. It was concluded that selection of the side-chain rotamers depends on the packing of  $\alpha$ -helices and, consequently, depends on the structural context.

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**Key words:**  $\alpha$ -proteins, rotamer preference, stereochemical analysis,  $\alpha$ -helix packing

## INTRODUCTION

As the number of resolved protein structures is increasing in the Protein Data Bank (PDB), an increasing number of works focus on side-chain rotamers in proteins. Most works are aimed at identification of various side-chain rotamers, their classification, statistical analysis of their frequencies in proteins, and construction of rotamer databases and libraries (e.g., [1–5]). Significant differences in side-chain rotamer frequencies have been observed as dependent on the local conformation of the main chain [1–7] and the membrane environment (for membrane proteins) [8]. These studies have shown that, in proteins, side chains adopt one of the three sterically possible rotamers, which are designated as  $g^-$  ( $\chi_1 = -60^\circ$ ),  $g^+$  ( $\chi_1 = +60^\circ$ ), and t ( $\chi_1 = 180^\circ$ ). In  $\alpha$ -helical regions, the main chain imposes additional limitations on the conformational freedom of side chains, and  $C_\beta$ -branched side chains contained in  $\alpha$ -helices have only one allowable conformer (Val, t; Ile and Trh,  $g^-$ ), while side chains nonbranched at  $C_\beta$  can have either  $g^-$  or t rotamers.

However, the other factors affecting the selection of side-chain conformations in proteins are still poorly understood. We have earlier shown that a close packing of hydrophobic side chains on the surfaces of  $\alpha$ -helices is reached owing to certain combinations of rotamers [9, 10]. The conformation of polar side chains can be selected depending on their structural environment. When a polar side chain has a partner to

form a hydrogen or salt bond, one (e.g., t) conformation is selected; otherwise, the side chain adopts another conformation so that its polar moiety is accessible for water molecules [10]. In this work, we considered the role of the structural context in selection of the conformation of hydrophobic side chains with the example of  $\alpha$ -helical leucine-zipper proteins. As a result, we observed new features of the distribution of  $g^-$  and t rotamers of side chains in hydrophobic cores of  $\alpha$ -helical structures and revealed a dependence of the side-chain conformation on the mode of  $\alpha$ -helix packing.

## EXPERIMENTAL

Our main subject was a class of coiled-coil (cc) proteins, also known as leucine zippers. Such proteins consist of long  $\alpha$ -helices, which are packed in a parallel or antiparallel manner to form dimers, trimers, tetramers, or, rarely, pentamers. These can be homo- or heterooligomers. In most known cc proteins,  $\alpha$ -helices are packed with angle  $\Omega \approx 20^\circ$ . In the ideal case, this angle between axes is characteristic of  $\alpha$ -helices whose amino acid sequences have heptad repeats (abcdefg)<sub>n</sub> with a- and d-positions occupied usually by hydrophobic residues (for a review, see [11, 12]).

Such proteins are best suited for our purpose for several causes. First, many cc proteins and their model analogs have Leu in the a- and d-positions. Such an almost homogeneous set of side chains makes the

packing of  $\alpha$ -helices nearly ideal, in contrast to heterogeneous sets, especially those with side chains dramatically differing in size. To study the selection of side-chain rotamers, our sample included only the cc proteins having the a- and d-positions occupied mainly by Leu, Phe, and Tyr residues (their side chains are most similar in size and can have two rotamers), but not proteins containing mostly Val and Ile in the a- and d-positions (see Tables 1–5). In addition, our database included homologous proteins and their model analogs that can be switched, e.g., from dimers to trimers or tetramers as a result of point mutations. This is of special importance for studying the effect of the structural context on the selection of side-chain conformations, because the same or a very close set of side chains occurs in different structural environments in such cases. Another advantage of cc proteins is that their  $\alpha$ -helices are rather long (mostly, of 20 or more residues), which substantially reduces the marginal effects. Necessary structural information was taken from PDB (<http://www.rcsb.org/pdb/>). Torsion angle  $\chi_1$  was computed using the MOLMOL program [13].

## RESULTS AND DISCUSSION

The packing of  $\alpha$ -helices against each other in proteins follows two main patterns, face-to-face and side-by-side, which differ in the arrangement of hydrophobic surfaces (or clusters) [9, 10, 12]. In the case of face-to-face packing, hydrophobic clusters are between the backbones of  $\alpha$ -helices in the contact region and form a bilayer of hydrophobic side chains. Such a packing is observed in all dimeric cc proteins with a parallel or antiparallel arrangement of  $\alpha$ -helices and in long isolated  $\alpha$ -helical hairpins of globular proteins (Tables 1, 2). In the case of side-by-side packing, hydrophobic clusters are combined to form a single layer of hydrophobic side chains on the surface of the double-helical structure. Such  $\alpha$ -helix packing is observed in tetramers and pentamers of cc proteins (Tables 4, 5) and in four-helix bundles of globular proteins. In trimeric cc proteins, the packing of  $\alpha$ -helices is most similar to side-by-side packing but shows some features of face-to-face packing (Table 3). Thus, depending on the  $\alpha$ -helix packing, hydrophobic side chains are arranged differently relative to each other and to the backbones of the  $\alpha$ -helices and, consequently, occur in different structural environments, which substantially affects their conformation.

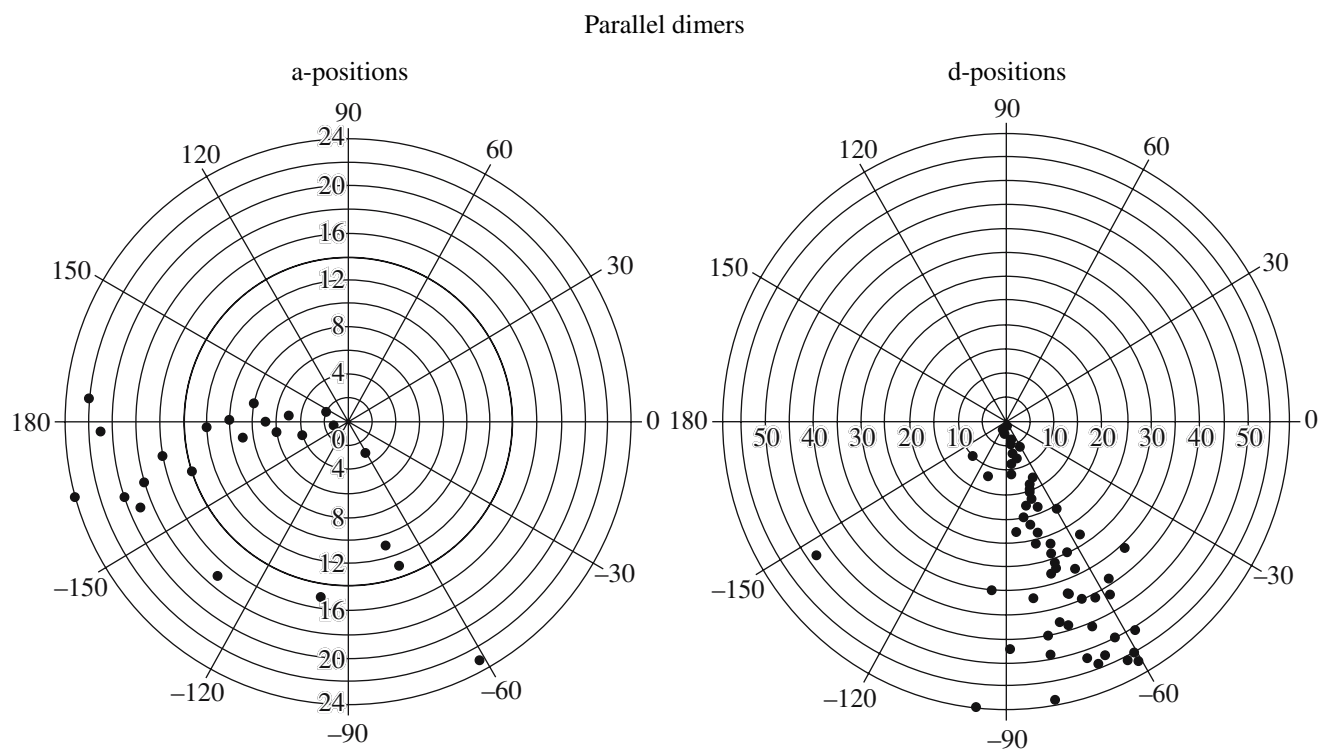
Table 1 summarizes the angles  $\chi_1$  computed for all residues occupying the a- and d-positions of cc dimers whose  $\alpha$ -helices are packed face-to-face and are parallel. Analysis of these data revealed an important feature in the distribution of rotamers of the hydrophobic side chains that are sterically allowed to adopt two conformations (Leu, Phe, Tyr, etc.). Most of the chains occur as t rotamers in the a-positions and as g<sup>-</sup> rotamers in the d-positions. To illustrate, Fig. 1 shows

a circular diagram of the  $\chi_1$  distribution for Leu residues occurring in the a- and d-positions of cc dimers with parallel  $\alpha$ -helices.

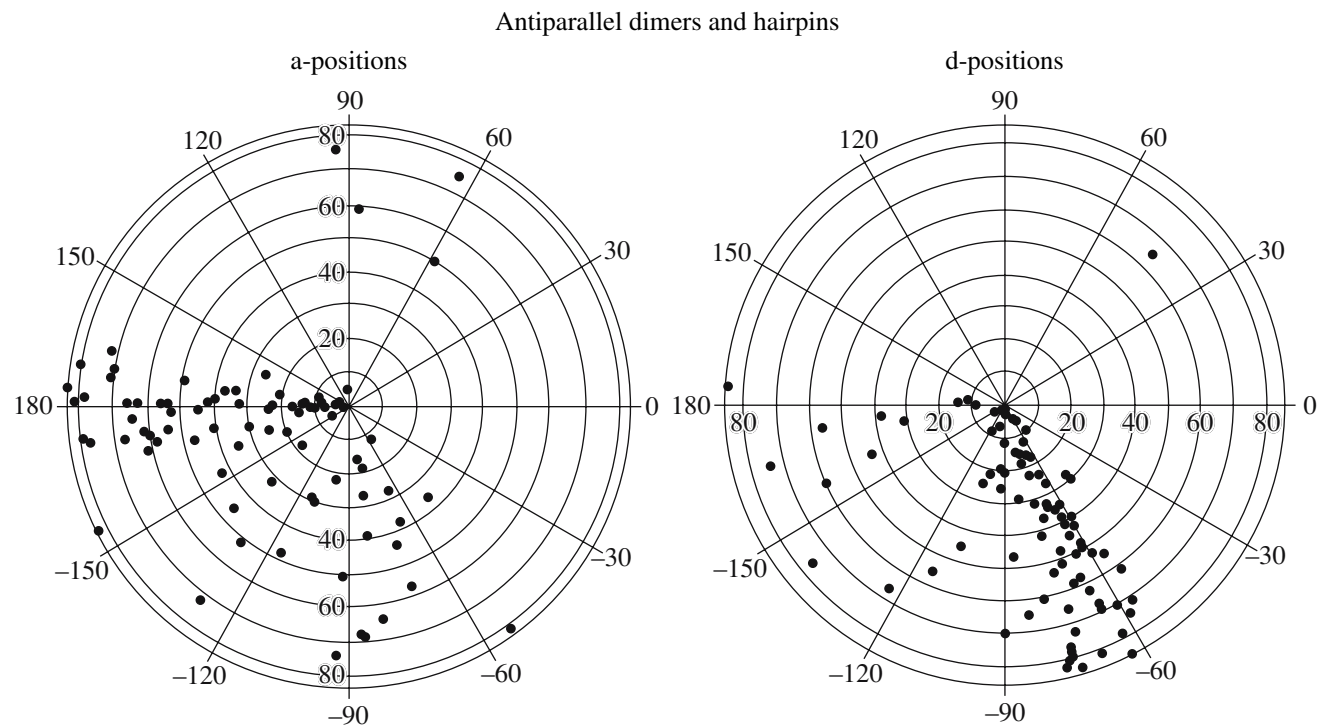
Angles  $\chi_1$  computed for residues occupying the a- and d-positions in  $\alpha$ - $\alpha$ -hairpins and antiparallel dimers are summarized in Table 2; their distribution is shown in Fig. 2. Although less distinct, the feature observed in this case was the same as in dimers with parallel  $\alpha$ -helices. The side-chain conformation will be considered a t rotamer when  $\chi_1$  ranges from  $-150^\circ$  to  $+150^\circ$  and a g<sup>-</sup> rotamer when  $\chi_1$  ranges from  $-50^\circ$  to  $-100^\circ$ . These criteria are more stringent than, for example, in [7], where  $\chi_1 = 180 \pm 60^\circ$  suggested t rotamers and  $\chi_1 = 0$  to  $-120^\circ$ , g<sup>-</sup> rotamers. A simple calculation shows that 64% of the side chains of Leu, Phe, and Tyr occupying the a-positions in antiparallel dimers and hairpins adopt t rotamers; 19% adopt g<sup>-</sup> rotamers, and 17% have sterically prohibited conformations (Fig. 2). For a comparison, 75% of Leu residues in the a-positions of dimers with parallel  $\alpha$ -helices adopt t rotamers, 21% adopt g<sup>-</sup> rotamers, and 4% have sterically prohibited conformations (Fig. 1). In the d-positions of hairpins and antiparallel dimers, 76% of Leu, Phe, and Tyr side chains adopt g<sup>-</sup> rotamers, 12% adopt t rotamers, and 12% have sterically prohibited conformations (Fig. 2). In parallel dimers, virtually all Leu residues in the d-positions adopt g<sup>-</sup> rotamers (Fig. 1). We think that the distribution (Fig. 2) displayed a greater number of deviations from the general trend as compared to that in Fig. 1 because the side-chain composition of antiparallel dimers and hairpins was more heterogeneous.

The distribution of side-chain rotamers in trimeric proteins substantially differed from that in dimers. As evident from Table 3 and Fig. 3, most (75%) of the a-position side chains of trimers adopt g<sup>-</sup> rotamers, rather than t rotamers, as in dimers. As for the d-positions, the situation with trimers was similar to that with dimers: 63% of the residues adopt g<sup>-</sup> rotamers, 25% adopt t rotamers, and 12% have sterically prohibited conformations. A similar distribution was observed for tetramers-1 (the a- and d-positions of tetramers-1 are occupied mostly by Leu, while the composition of tetramers-2 is heterogeneous): 82% of Leu residues in the A-positions adopt g<sup>-</sup> rotamers, t rotamers are absent, and 18% of residues have sterically prohibited conformations (Table 4, Fig. 4). Of the five residues found in the d-positions of tetramers-1, three adopt g<sup>-</sup> rotamers and two adopt t rotamers.

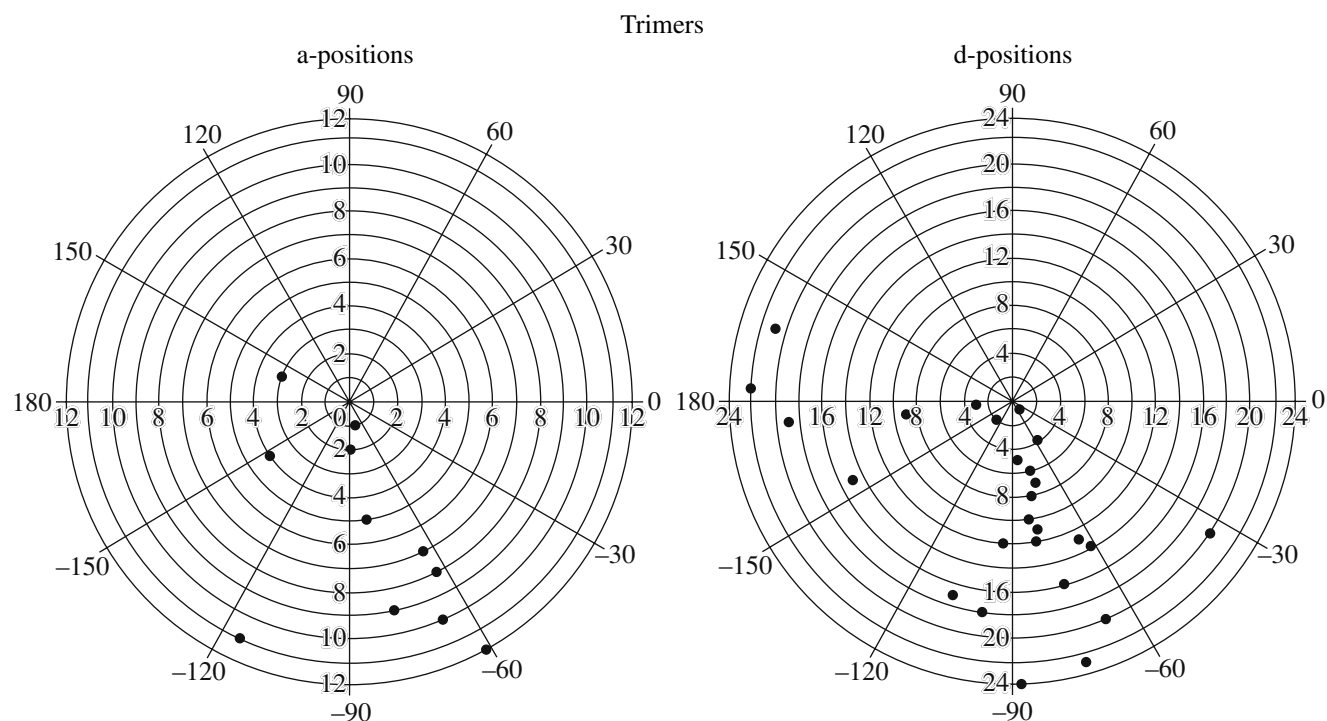
The rotamer distribution in tetramers-2 is opposite to that in dimers: most side chains in the a-positions adopt g<sup>-</sup> rotamers, while side chains in the d-positions mostly adopt t rotamers (Table 5, Fig. 5). According to our estimates, 83% of the Leu, Phe, and Tyr residues in the a-positions of tetramers-2 adopt g<sup>-</sup> rotamers, 8% adopt t rotamers, and 9% have sterically prohibited conformations. In the d-positions of tetramers-2,



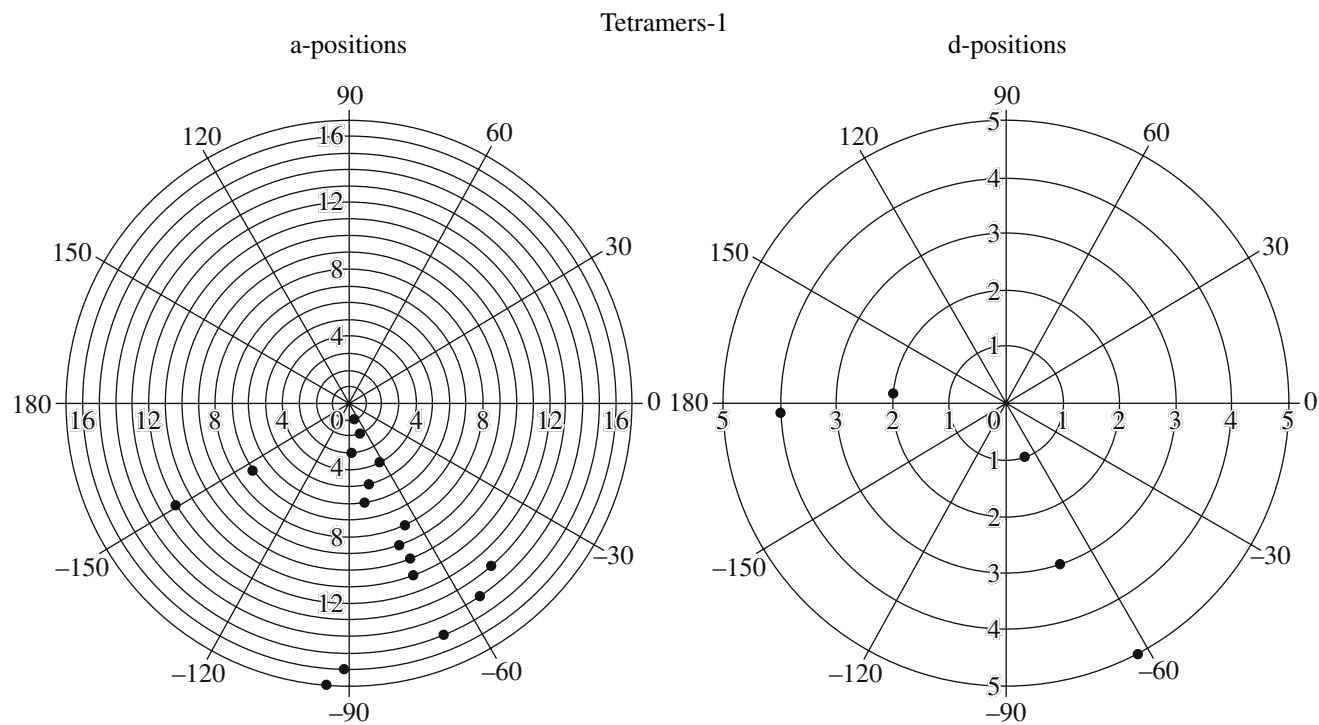
**Fig. 1.** Torsion angle  $\chi_1$  distribution of Leu residues occupying the a- and d-positions in parallel cc dimers (based on Table 1). Here and in Figs. 2–5, distributions are shown as circular diagrams. The upper half-circles correspond to positive ( $\chi_1$  ranging from 0 to 180°) angles and the lower ones, to negative ( $\chi_1$  ranging from 0 to –180°) angles.



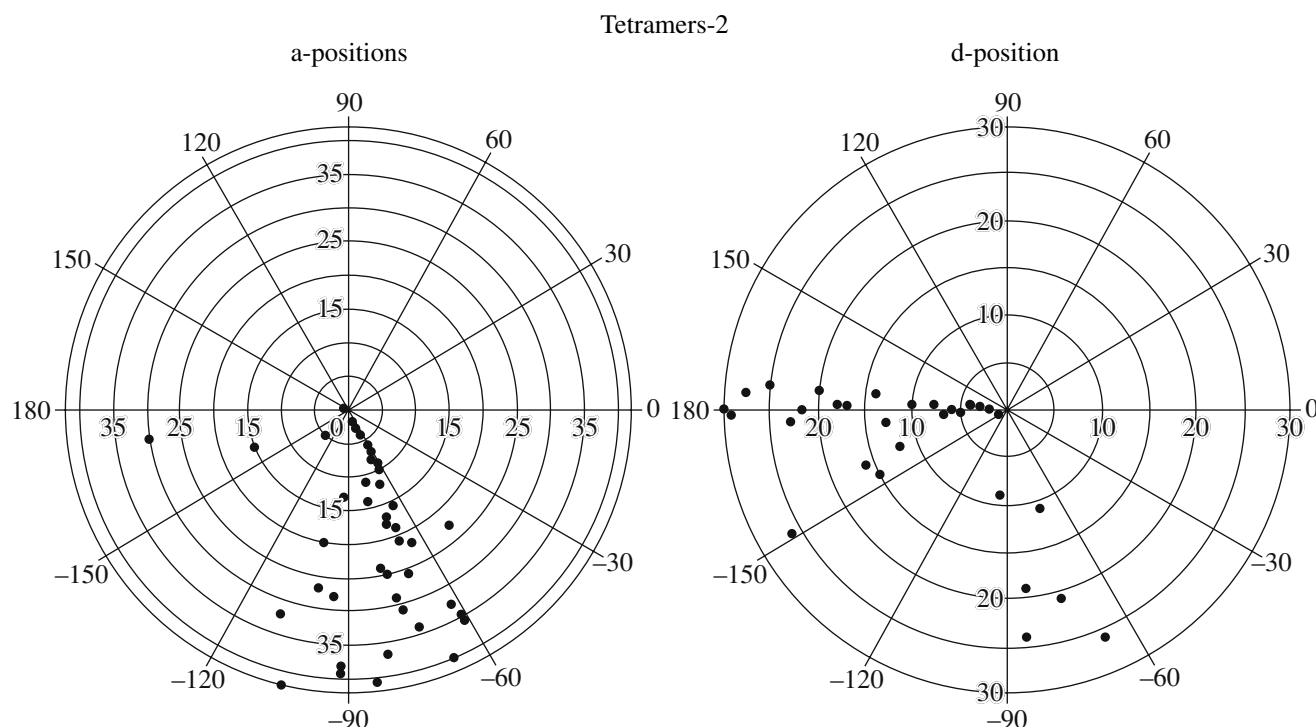
**Fig. 2.** Torsion angle  $\chi_1$  distribution of Leu, Phe, and Tyr residues occupying the a- and d-positions in antiparallel cc dimers and  $\alpha$ -helical hairpins (based on Table 2).



**Fig. 3.** Torsion angle  $\chi_1$  distribution of Leu, Phe, and Tyr residues occupying the a- and d-positions in trimers (based on Table 3).



**Fig. 4.** Torsion angle  $\chi_1$  distribution of Leu residues occupying the a- and d-positions in tetramers-1 (based on Table 4).



**Fig. 5.** Torsion angle  $\chi_1$  distribution of Leu, Phe, and Tyr residues occupying the a- and d-positions in tetramers-2 (based on Table 5).

80% of residues adopt t rotamers and 20% adopt  $g^-$  rotamers.

Thus, a trend is distinct: the conformation of side chains changes from t to  $g^-$  rotamers in the a-positions and from  $g^-$  to t rotamers in the d-positions upon the transition from dimers to trimers and tetramers; i.e., as the  $\alpha$ -helix packing changes from face-to-face to side-by-side. In the intermediate cases of trimers and tetramers-1, especially with a homogeneous set of side chains,  $g^-$  rotamers are prevalent in both a- and d-positions. In other words, the selection of side-chain rotamers in the a- and d-positions depends on the  $\alpha$ -helix packing and, consequently, on the structural context.

The trend observed can be explained by a simple squeezing mechanism; i.e., the side chains are squeezed out of the helix-helix interface. A possible scenario is considered below with the example of dimers. Assume that dimers initially have  $g^-$  rotamers of side chains in the a-positions and t rotamers in the d-positions. In this case, a major part of each side chain in these positions occur in the space between the backbones of  $\alpha$ -helices. As the a-position side chains adopt t rotamers and the d-position side chains adopt  $g^-$  rotamers (as observed), the centers of mass of the side chains are displaced from the helix-helix interface outwards, and the tendency to a closer packing reduces the distance between the axes of the  $\alpha$ -helices. Thus, the interactions between helices and the tendency to a closer packing bring the axes of the  $\alpha$ -

helices close together and displace the side chains from the between-helix space. This mechanism will be detailed elsewhere.

The trends observed in the rotamer distribution in dimers, trimers, and tetramers of cc proteins are of importance. Advantage of them can be taken in modeling protein structures and designing new structures in protein engineering. It should be noted that the side-chain rotamers can be predicted not only for Leu, Phe, and Tyr, but also for  $\beta$ -branched Val and Ile, because these residues each have a single sterically allowable rotamer in the vast majority of cases (as can be seen from Tables 1–5).

The observed deviations from the above trends can be explained by the effects of the following factors. Analysis showed that such deviation occur mostly at the ends of helices; i.e., marginal effects do take place. This is possibly because  $\alpha$ -helices are diverged from each other at the ends of some oligomers and no longer affect each other. The heterogeneity of the sets of a- and d-position side chains can further contribute to the deviations. For instance, the trends are more distinct in parallel dimers, which have almost homogeneous sets of Leu residues (Fig. 1), than in antiparallel dimers, whose sets of side chains are heterogeneous (Fig. 2).

To conclude, it should be noted that similar trends in the distributions of side-chain rotamers were observed for four-helix bundles (our preliminary data)

**Table 1.** Angle  $\chi_1$  for residues in the **a**- and **d**-positions in dimers of cc proteins with a parallel  $\alpha$ -helix packing\*

No.	PDB ID	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d
1	1D7M(A)		M244	L248	L251	L255	E258	R262	L265	K269	L272	L276	L279	G283	R286	L290	L293	L297	T300	N304	E307
			-72	-164	-84	-172	-84	171	-73	-179	-89	-98	-69	-	171	-162	-80	-178	-56	-61	-142
		R311	L314	L318	T321	R325	L328	L332	A335	K339	L342										
		169	-78	174	-67	179	-85	-178	-	-164	-145										
2	1DH3(A)	E287	L290	R294	A297	R301	K304	V308	L311	V315	L318	N322	L325	L329	L332						
		-155	-103	102	-	-136	-120	-26	-65	95	-79	-93	-61	-131	-61						
3	1GD2(E)	L101	L104	V108	L111	H115	T118	N122	L125	V129	L132	L136	L139								
		-167	-92	169	-109	169	50	174	-95	79	-63	-170	-								
4	1ZII(A)	M2	L5	V9	L12	A16	L19	V23	L26	V30	R33										
		-176	-77	167	-65	-	-74	171	-47	164	-										
5	1UIX(A)	V982	L985	K989	L992	L996	A999	L1003	L1006	E1010	A1013	K1017	F1020	L1024	E1027	K1031	A1034	L1038			
		171	-78	-72	-69	179	-	-71	-81	-	-	162	-96	-164	-76	171	-	175			
6	1KDD(A)	V2	L5	V9	L12	L16	L19	V23	L26	N30	C33										
		178	-63	167	-70	-73	-74	166	-70	-173	-177										
7	1CZ7(A)	L321	C324	L328	S331	R335	L338	V342	L345												
		157	180	-172	-165	-169	-76	162	-57												
8	1KQL(A)	V32	L35	N39	L42	V46	L49	V53	L56	L60	Q63	Y67	I70	L74	A77	M81					
		171	-78	-82	-71	167	-70	162	-67	-163	-164	-165	-133	-158	-	172					
9	1S9K(D)		L165	T169	L172	K176	L179	I183	L186	K190											
			-74	56	-72	-178	-57	-66	-63	170											
10	1S9K(E)	R280	K283	I287	L290	V294	L297	N301	L304	A308	L311	V315	L318								
		-83	-159	-53	-76	-175	-71	-89	-59	-	-58	168	-63								
11	1LLM(C)	H50	I53	L57	L60	V64	L67	N71	L74	V78	L81	V85									
		159	-59	-180	-70	169	-65	-75	-75	166	-71	-39									
12	1CE9(A)	V3	L6	V10	L13	N17	L20	V24	L27	V31	R34										
		47	-83	167	-80	-84	-71	165	-73	149	-104										
13	1T6F(A)	L2	A5	N9	L12	I16	K19	I23	L26	N30	L33	A37									
		-104	-	-172	-60	-68	-68	-69	-79	-72	-67	-									
14	1P9I(A)	M1	L4	L8	L11	N15	L18	V22	L25	V29											
		-	-135	169	-77	174	-71	166	-67	163											

Note: \*Here and in Tables 2–5, angle  $\chi_1$  is measured in degrees and is indicated below the corresponding residue.

**Table 2.** Angle  $\chi_1$  for residues in the **a**- and **d**-positions in dimers of cc proteins and  $\alpha$ -helical hairpins with an antiparallel and face-to-face  $\alpha$ -helix packing

No.	PDB ID	a	d	a	d	a	d	a	d	a	d	a	d	a	d
1	1A32(1)		P24	I28	L31	I35	L38	L42	L38	L42	L38	L42	L38	L42	L38
2	1A32(2)		–	–65	–88	–67	–171	–92	–171	–92	–171	–92	–171	–92	–171
3	1A36(1)	M644	S51	L55	M58	R62	L65	L69	L65	L69	L65	L69	L65	L69	L65
4	1A36(2)	–	–57	177	–73	–170	–74	–129	–74	–129	–74	–129	–74	–129	–74
5	1AQT(1)	E91	L647	I651	K654	L658	A661	L665	A661	L665	A661	L665	A661	L665	A661
6	1AQT(2)	–	–74	–67	–35	173	–	–178	–	–178	–	–178	–	–178	–
7	1CXZ(1)	–	T680	V684	K687	V691	L694	L698	L694	L698	L694	L698	L694	L698	L694
8	1CXZ(2)	–	–56	179	–76	172	–68	179	–68	179	–68	179	–68	179	–68
9	1IDG3(1)	–	A28	Q32	L35	R39	L42	I46	L42	I46	L42	I46	L42	I46	L42
10	1IDG3(2)	–	V73	L77	S80	L84	L87	L91	L87	L91	L87	L91	L87	L91	L87
11	1IE79(1)	–	–152	–81	–175	–82	–62	–173	–62	–173	–62	–173	–62	–173	–62
12	1IE79(2)	–	R539	L543	Q546	L550	K553	Q557	K553	Q557	K553	Q557	K553	Q557	K553
13	1IFXK(A1)	–	–175	–180	–80	177	–74	–168	–74	–168	–74	–168	–74	–168	–74
14	1IFXK(A2)	–	M572	I576	L579	M583	L579	L579	L579	L579	L579	L579	L579	L579	L579
15	1IFXK(A3)	–	–77	–65	177	–70	–70	–70	–70	–70	–70	–70	–70	–70	–70
16	1IFXK(A4)	–	A108	L112	A115	L119	L119	L119	L119	L119	L119	L119	L119	L119	L119
17	1IGRJ(1)	–	–	–141	–	–171	L145	L145	L145	L145	L145	L145	L145	L145	L145
18	1IGRJ(2)	–	–41	–66	–75	–176	–65	–65	–65	–65	–65	–65	–65	–65	–65
19	1IGRJ(2)	–	Q10	F14	L17	A21	I24	K28	I24	K28	I24	K28	I24	K28	I24
20	1IGRJ(2)	–	–95	179	–79	–	–78	–166	–78	–166	–78	–166	–78	–166	–78
21	1IGRJ(2)	–	K67	T71	L74	L78	L81	E85	L81	E85	L81	E85	L81	E85	L81
22	1IGRJ(2)	–	165	–61	–74	178	–72	174	–72	174	–72	174	–72	174	–72
23	1IGRJ(2)	–	I10	L14	Y17	V21	I24	M28	I24	M28	I24	M28	I24	M28	I24
24	1IGRJ(2)	–	–55	–77	–75	168	–65	–73	–65	–73	–65	–73	–65	–73	–65
25	1IGRJ(2)	–	I97	K101	L104	L108	M111	L115	M111	L115	M111	L115	M111	L115	M111
26	1IGRJ(2)	–	–67	–64	–67	–153	–69	179	–69	179	–69	179	–69	179	–69
27	1IGRJ(2)	–	L14	L18	L21	R26	I29	I33	I29	I33	I29	I33	I29	I33	I29
28	1IGRJ(2)	–	–122	–158	–57	176	–72	–51	–72	–51	–72	–51	–72	–51	–72
29	1IGRJ(2)	–	A51	Q55	C58	I62	I65	L69	I65	L69	I65	L69	I65	L69	I65
30	1IGRJ(2)	–	–	–146	–68	–68	–64	–71	–64	–71	–64	–71	–64	–71	–64
31	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
32	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
33	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
34	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
35	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
36	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
37	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
38	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
39	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
40	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
41	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
42	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
43	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
44	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
45	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
46	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
47	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
48	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
49	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
50	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
51	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
52	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
53	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
54	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
55	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
56	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
57	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
58	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
59	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
60	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
61	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
62	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
63	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
64	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
65	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
66	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
67	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
68	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
69	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
70	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
71	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
72	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
73	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
74	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
75	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
76	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
77	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
78	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
79	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
80	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
81	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
82	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
83	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
84	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
85	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
86	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
87	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
88	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
89	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
90	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
91	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
92	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
93	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
94	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
95	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
96	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
97	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
98	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
99	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
100	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
101	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
102	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
103	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
104	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
105	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
106	1IGRJ(2)	–	–	–	–	–	–	–	–	–	–	–	–	–	–
107	1IGRJ(2)	–	–												





Table 2. (Contd.)

No.	PDB ID	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d
17	1A92(A)	R13	I16	W20	G23	L27	L30	L34	L37	I41	L44						
		-74	-63	-173	-	-168	-75	-168	-173	-64	-123						
18	1GMJ(A)	I53	H56	I60	L63	I67	H70	I74	L77	E81							
		-75	-74	-74	-65	-70	-73	-71	-67	-161							
19	1Q05(A)	K81	T84	V88	I91	I95	L98	R102	L105	A109	C112						
		-174	-54	165	-62	-68	-61	-170	-69	-	171						
20	1YF2(A)		I174	L178	I181	I185	I188	I192	L195	K199	L202						
			-65	-65	-56	-66	-68	-71	-157	168	-65						
21	2C2A(A)	L285	L288	L292	I295	S299	L302	L306	L309								
			-115	-112	-71	-76	-160	179	174								
22	2AYU(A)	V92	K95	L99	L102	L106	V109	F113	E116	L120	K123						
		174	-55	-110	-107	-115	-166	173	-69	-172	172						
23	1ZHC(A)	F19	I22	H26	L29	I33	A36	N40									
		95	-49	177	-137	46	-159	-90									
		V46	M49	K53	L56	I60	M63	Y67	K70								
		-168	-61	-154	-94	-75	-162	-154	73								
24	1K1F(A)	V28	I31	L35	A38	I42	L45	V49	E52	M56	L59						
		168	-43	-169	-	-74	-64	170	-63	-	-65						
25	1ENV(A)		A30	L34	I37	Q41	L44	I48	Q51	L55	T58						
			-	-136	-74	-59	-108	-73	-76	-87	62						
		W117	W120	I124	Y127	I131	L134	S138	Q141	N145	E148						
		-171	-177	-87	-77	-58	-87	-64	-103	-66	-77						
26	1JAL(A)		A133	L137	C140	I144	L147	A151	G154								
			-	178	179	-79	-70	-	-								
		L162	M165	L169	L172												
		176	-64	172	-63												
27	1QVR(A)	I403	L406	K410	L413	R417	L420										
		-80	-54	-179	-50	70	-62										
			I435	I439	L442	I446	L449	W453	E456								
			-179	-79	-82	-69	-62	-179	-89								
			I459	L463	A466	L470	V473	I477	A480	Y484	N487						
			-61	-161	-	171	167	-72	-	176	-49						
			R493	L497	L500	V504	L507	L511									
			-174	172	-49	94	-59	-65									

**Table 3.** Angle  $\chi_1$  for residues in the **a**- and **d**-positions of trimers

No.	PDB ID	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>
1	1AQ5(A)	Q18 -88	V21 154	I25 -148	L28 -84	L32 160	V35 142	I39 -80	L42 -74
2	1B08(B)	V1204 -	L1207 -49	V1211 177	L1214 -75	V1218 175	L1221 -79	F1225 -158	Y1228 -88
3	1EQ7(A)		S2 65	I6 -72	L9 -74	V13 167	L16 -80	V20 166	L23 -107
		A41 -	A44 -	L48 -78	M51 60	Y55 -61			
4	1SWI(A)	M2 -49	L5 -128	V9 175	L12 -78	A16 -	L19 -64	V23 147	L26 -98
5	2BEZ(C)	Y899 -	Q902 -	A906 -	F909 178	I913 -82	I916 -77	L920 -64	T923 -64
			V934 162	A938 -	L941 -173	V945 174	L948 -61	F952 -75	I955 -68
6	1G2C(A)		L160 -174	V164 172	I167 -74	L171 -146	T174 -72	V178 171	L181 -175
7	1COS(B)	W2 -173	L5 -56	L9 -90	L12 -82	L16 -82	L19 -154	L23 -63	L26 -34
	1COS(C)	-60	-83	-115	-65	-94	-80	-78	163
No.	PDB ID	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>
1	1AQ5(A)	I46 -68							
2	1B08(B)	E1232 -133							
3	1EQ7(A)	V27 170	M30 -140	V34 178	A37 -				
4	1SWI(A)	V30 141							
5	2BEZ(C)								
		L959 -67	I962 -71	L966 -115	V969 156				
6	1G2C(A)	V185 168	L188 177	V192 165	L195 -74	I199 -64	Q202 -62	I206 67	K209 -
7	1COS(B)								

**Table 4.** Angle  $\chi_1$  for residues in the **a**- and **d**-positions of tetramers-1

No.	PDB ID	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>	<b>a</b>	<b>d</b>
1	2BNI(A)	M2 -59	I5 -69	L9 -77	I12 -70	G16 -	I19 -69	L23 -69	I26 -77	L30 -56	
2	2B1F(B)	V2 -69	L5 -70	V9 169	L12 -71	N16 -151	L19 -178	V23 165	L26 -62	V30 167	
3	1W5K(A)	M3 -170	I6 -68	L10 -73	I13 -77	L17 -82	I20 -72	L24 -70	I27 -73	L31 -68	
4	1C94(A)	R4 57	V7 168	L11 -88	V14 162	L18 -146	N21 -77	L25 -150	V28 169	L32 -91	M35 49
5	4HB1(A)		L4 175	L8 -63	A11 -	L15 -66	A18 -	A22 -			
6	1GCL(A)	M2 -59	I5 -87	L9 -77	I12 -58	L16 -71	I19 -82	L23 -49	I26 -67	L30 -95	

**Table 5.** Angle  $\chi_1$  for residues in the **a**- and **d**-positions of tetramers-2

No.	PDB ID	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d	a	d
1	ISFC(A)	L32 175	T35 -53 A191	V39 177 I195	V42 -173 R198	M46 -67 I202	N49 -79 L205	V53 109 I209	R56 169 L212	L60 -79 F216	L63 178 M219	A67 - V223	L70 -177 Q226	A74 - I230	F77 -179 I233	A81 - V23	L84 -151 A240	Y88 -61	K91 170		
	ISFC(B)		-	I195	R198	I202	L205	I209	L212	F216	M219	V223	Q226	I230	I233	V23	A240				
	ISFC(C)		S25 -75 T46	T29 57 L50	M32 -76 Q53	V36 89 L57	S39 -46 V60	G43 - M64	-174 - I67	-84 -92 M71	-82 - A74	-22 -76 L78	-47 -85 L81	-69 - A185	-64 -65 N188	173 S61 I192	-		M202 -57		
	ISFC(D)		M146 -76 L15	L150 -62 V19	V153 178 V22	I157 -60 M26	L160 -95 N29	A164 - I33	-64 -173 R36	-92 -54 L40	- Q174 L43	I178 -70 T47	I181 -70 L50	A185 - S54	N188 -65 F57	-76 S61 I192	A195 V64 A220	A199 - L224	M202 -57		
2	IGL2(A)	V12 175	L15 -152	V19 169	V22 178	M26 -60	N29 -95	I33 - R36	L40 -54	L40 -54	L43 -65	T47 -70	L50 -70	S54 70	F57 -180	S61 -67	V64 64		A227 -		
	IGL2(B)	M168	R171	I175	L178	I182	I185	F189	L192	I196	Q199	I203	I206	V210	A213	V217	A220	L224	A227		
	IGL2(C)	-	S149	A153	T156	G160	I163	L167	Q170	L174	T177	L18	T18	L188	S191	L195	-	-	-		
	IGL2(D)	L155	L158	I162	Q165	G169	I172	L176	Q179	I183	L186	V19	T19	L197	E200	-	-	-	-		
3	IHV(V)(D)	I195	R198	I202	L205	I209	L212	F216	M219	V223	Q226	I230	I233	V237	A240	V244	A247				
4	IVDF(A)*	-78	177	-69	-180	-70	177	-67	179	171	-64	-69	-74	168	-	-56	-				
5	INHL	A36	S39	G43	T46	L50	Q53	L57	I60	L64	I67	M71	T74	L78	L81	-	-				
6	IJTH(A)	-	173	-	-57	-79	-159	-69	-70	-100	-65	-	-70	-62	-67	-	-				
	IJTH(A)	D4	M7	L11	M14	A18	L21	S25	S28	M32	L35	S39	A42	-	-	-	-				
	IJTH(A)	L50	Q53	L57	I60	M64	I67	M71	A74	L78	L81	-	-	-	-	-	-				
	IJTH(B)	I195	R198	I202	L205	I209	L212	F216	M219	V223	Q226	I230	I233	V237	A240	V244	A247				
7	2B1F(D)	L5	A8	L12	A15	L19	A22	L26	L29	M32	L35	S39	A42	-	-	-	-				
8	4HB1(A)	-67	-	-62	-	-160	-	-65	-154	-	-	-	-	-	-	-	-				
9	1GCL(A)	M2	I5	L9	I12	L16	I19	L23	I26	L30	-	I230	I233	V237	A240	V244	A247				
	1GCL(A)	-59	-87	-77	-58	-71	-82	-49	-67	-95	-	-78	-73	171	-	-	-				
10	1GI1(A)	I95	I95	M99	V102	M106	Q109	I113	L116	E120	-	-	-	-	-	-	-				
	1GI1(A)	-	48	-85	161	-77	-69	-65	-159	-66	-	-	-	-	-	-	-				
11	1HTM(B)**	N49	L52	I56	T59	F63	I66	F70	V73	I77	L80	V84	T87	L91	Y94	V237	-				
	1HTM(B)**	-88	174	-62	66	81	-63	-92	147	-64	180	168	-75	-77	176	-	-				

Notes: \* Pentamer.  
\*\* Trimer.

and other  $\alpha$ -helical proteins (see [9, 10]), which are a subject of our further studies.

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### REFERENCES

1. Janin J., Wodak S., Levitt M., Maigret B. 1978. Conformation of amino acid side-chains in proteins. *J. Mol. Biol.* **125**, 357–386.
2. Ponder J.W., Richards F.M. 1987. Tertiary templates for proteins. Use of packing criteria in the enumeration of allowed sequences for different structural classes. *J. Mol. Biol.* **193**, 775–791.
3. Schrauber H., Eisenhaber F., Argos P. 1993. Rotamers: To be or not to be? An analysis of amino acid side-chain conformations in globular proteins. *J. Mol. Biol.* **230**, 592–612.
4. Dunbrack R.L., Jr., Karplus M. 1993. Backbone-dependent rotamer library for proteins: Application to side-chain prediction. *J. Mol. Biol.* **230**, 543–571.
5. Lovell S.C., Word J.M., Richardson J.S., Richardson D.C. 2000. The penultimate rotamer library. *Proteins*. **40**, 389–408.
6. McGregor M.J., Islam S.A., Sternberg M.J.E. 1987. Analysis of the relationship between side-chain conformation and secondary structure in globular proteins. *J. Mol. Biol.* **198**, 295–310.
7. Dunbrack R.L., Jr., Cohen F.E. 1997. Bayesian statistical analysis of protein side-chain rotamer preferences. *Protein Sci.* **6**, 1661–1681.
8. Chamberlain A.K., Bowie J.U. 2004. Analysis of side-chain rotamers in transmembrane proteins. *Biophys. J.* **87**, 3460–3469.
9. Efimov A.V. 1977. Stereochemistry of  $\alpha$ -helix and  $\beta$ -structure packing in a compact globule. *Dokl. Akad. Nauk SSSR*. **235**, 699–702.
10. Efimov A.V. 1979. Packing of  $\alpha$ -helices in globular proteins. Layer-structure of globin hydrophobic cores. *J. Mol. Biol.* **134**, 23–40.
11. Lupas A. 1996. Coiled coils: New structures and new functions. *Trends Biochem. Sci.* **21**, 375–382.
12. Efimov A.V. 1999. Complementary packing of  $\alpha$ -helices in proteins. *FEBS Lett.* **463**, 3–6.
13. Koradi R., Billeter M., Wutrich K. 1996. MOLMOL: A program for display and analysis of macromolecular structures. *J. Mol. Graph.* **14**, 51–55.